

INVITED REVIEWS AND SYNTHESSES

Isolation by environment

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Abstract

The interactions between organisms and their environments can shape distributions of spatial genetic variation, resulting in patterns of isolation by environment (IBE) in which genetic and environmental distances are positively correlated, independent of geographic distance. IBE represents one of the most important patterns that results from the ways in which landscape heterogeneity influences gene flow and population connectivity, but it has only recently been examined in studies of ecological and landscape genetics. Nevertheless, the study of IBE presents valuable opportunities to investigate how spatial heterogeneity in ecological processes, agents of selection and environmental variables contributes to genetic divergence in nature. New and increasingly sophisticated studies of IBE in natural systems are poised to make significant contributions to our understanding of the role of ecology in genetic divergence and of modes of differentiation both within and between species. Here, we describe the underlying ecological processes that can generate patterns of IBE, examine its implications for a wide variety of disciplines and outline several areas of future research that can answer pressing questions about the ecological basis of genetic diversity.

Keywords: gene flow, isolation by distance, isolation by environment, landscape genetics, population genetic structure

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Introduction

Seventy-one years ago, Sewall Wright (Wright 1943) introduced the term 'isolation by distance' (IBD) to describe a pattern in which genetic differentiation increases with geographic distance. The theory of isolation by distance describes the local accumulation of genetic differences when dispersal between populations or subgroups is geographically restricted (Slatkin 1993). Effectively, genetic differentiation between populations is the result of drift acting within populations more quickly than it is ameliorated by gene flow between populations (Slatkin 1993; Rousset 1997). Therefore, any processes that reduce the effective dispersal rate between populations will generate patterns of greater genetic differentiation (Slatkin 1993; Bolnick & Otto 2013).

Decades of investigation into IBD have revealed it to be common in nature (Slatkin 1993; Meirmans 2012) and brought a focus to the geography of population divergence and isolation (Mayr 1963). However, geography represents only one of the key landscape components that can potentially influence gene flow and population connectivity (Crispo *et al.* 2006; Lee & Mitchell-Olds 2011). Another important part of a landscape is the environment (Nosil *et al.* 2005; Foll & Gaggiotti 2006; Thorpe *et al.* 2008), and in the past decade, new fields such as landscape genetics (Storfer *et al.* 2007; Balkenhol *et al.* 2009; Wagner & Fortin 2013) have arisen to examine the roles played by ecology and the environment in micro-evolutionary processes (McRae & Beier 2007; Räsänen & Hendry 2008; Bolnick & Otto 2013). One of the important concepts that has emerged is 'isolation by environment' (IBE; Wang & Summers 2010), which encapsulates the relationship between environmental heterogeneity and spatial variation in gene flow on a landscape (Bradburd *et al.* 2013; Wang *et al.* 2013; Sexton *et al.* 2014).

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We define isolation by environment as a pattern in which genetic differentiation increases with environmental differences, independent of geographic distance, and which is agnostic with respect to the underlying processes that generated it. The study of IBE comprises two challenges. The first is to disentangle the relative strengths of IBD and IBE in observed patterns of spatial genetic differentiation, which can be a difficult statistical problem because geographic distance and environmental differences are often correlated (Fig. 1; Lee & Mitchell-Olds 2011; Wang 2013; Shafer & Wolf 2013; Bradburd *et al.* 2013). The second is then to determine the processes that have generated and maintained those patterns. The development of new methods in spatial statistics and the rapid proliferation of both genomic and environmental GIS data have greatly facilitated research to overcome the first challenge of quantifying IBE relative to IBD, but researchers seeking to tease apart these patterns must still take care to employ appropriate study design, sampling strategies and statistical techniques (Bradburd *et al.* 2013; Sexton *et al.* 2014). The second challenge requires assessing the ways in which drift, selection and dispersal have acted to shape patterns of genetic variation. Quantifying the relative contributions of these processes, the ways in which they depend upon an ecologically heterogeneous landscape, and how those relative contributions vary across species or landscapes is an active and exciting field of research.

Here, we describe the processes that can generate this pattern and discuss important methods and considerations for studying IBE. We conclude by proposing avenues for future research in IBE and suggesting a range of new and exciting questions about the ecological basis of spatial genetic variation. The rise of IBE as a research focus has resulted in a tremendous opportunity to examine, often at very fine scales, the ways in which ecology shapes genetic variation in nature, forming a

true bridge between the fields of population genetics and landscape ecology.

Definition of IBE

Isolation by environment is defined as a pattern in which genetic differentiation increases with environmental differences, independent of geographic distance (Fig. 1; Wang & Summers 2010; Bradburd *et al.* 2013; Sexton *et al.* 2014). The important environmental variables may be continuous, such as elevation or humidity (Murphy *et al.* 2010; Bradburd *et al.* 2013), or discrete, such as habitat or substrate type (Andrew *et al.* 2012). They may describe abiotic factors, such as temperature and precipitation (Wang 2012), or biotic factors, such as vegetation density and host (Via & Hawthorne 2002). The key is that they contribute to explaining variation in pairwise genetic distances beyond that explained by geographic distance (and after accounting for any correlation between environmental and geographic distances). This definition is solely a description of a pattern and is agnostic with respect to the processes that have generated that pattern.

We advocate for this pattern-based, rather than process-based, definition because there are frequently many biological processes that can generate a given pattern of observed genetic differentiation. This definition provides the general case for which numerous other 'isolation by' terms present specific, process-based instances. For example, isolation by adaptation (IBA), defined as the correlation of adaptive phenotypic and neutral genetic divergence (Nosil *et al.* 2008), and isolation by ecology, defined as the correlation of ecological and neutral genetic divergence (Claremont *et al.* 2011; Edelaar *et al.* 2012), both describe patterns due to a specific selective mechanism. These process-based terms are designed to address the important question of how observed patterns of spatial genetic variation are gener-

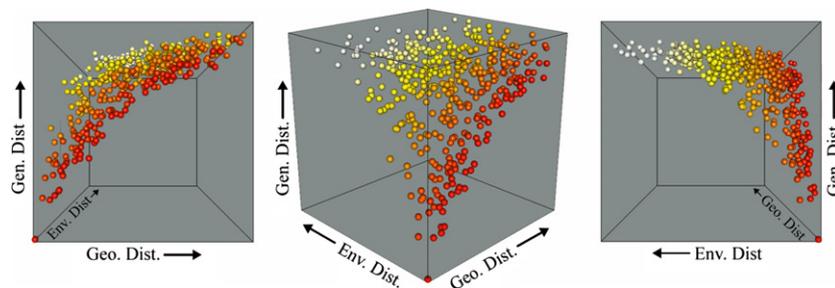


Fig. 1 Isolation by distance and environment. Under the patterns of isolation by distance (IBD) and isolation by environment (IBE), genetic distance increases with geographic and environmental distance. The three panels show different views of a simulated data set in which both patterns can be seen. Points represent the genetic distance (Gen. Dist.) between a pair of populations plotted against their geographic (Geo. Dist.) and environmental distances (Env. Dist.) and are heat-coloured by the magnitude of that environmental distance.

ated; however, the processes that have generated these empirical patterns of divergence cannot be observed directly from these patterns, and the same empirical pattern of genetic differentiation could be due to many different underlying processes.

Our definition can also be contrasted with isolation by resistance (IBR), defined as the correlation of genetic and 'resistance' distances (McRae & Beier 2007). Resistance distance between a pair of populations can be understood as the probability that an individual disperses from one to the other, integrating over all paths that individual might take, and weighting those paths by their 'friction' to dispersal (a low pairwise resistance means a high probability of dispersal, and vice versa). Isolation by resistance, like our definition of IBE, is also agnostic with respect to process; the dispersal resistance of a specific landscape element may be due to selection against maladapted dispersers, migratory preference or simple cost of transport. However, IBR implicitly conflates IBD and IBE, making it impossible to differentiate the strengths of these two patterns in empirical data.

Our simple but broad definition is meant to be inclusive of all positive associations between genetic and environmental distances, in keeping with the tradition set up by Wright's (1943) definition of IBD. It provides a collective term for a genetic pattern that is common in nature but may be generated by many different processes, and it implicitly links environmental variation on a landscape to gene flow and population structure. Thus, IBE is valuable for understanding spatial genetic differentiation in the context of landscape variation.

Processes generating IBE

Isolation by environment is a pattern that can be generated by a variety of ecological processes. These may be relatively simple, like when a temperature cline regulates dispersal among populations of an ectotherm (e.g. Murphy *et al.* 2010), or they may represent more complex ecological interactions, like when genetic differentiation between plant populations is mediated by differences in their pollinator communities (e.g. Hopkins *et al.* 2012). This many-to-one mapping of process to pattern can make it difficult, though by no means impossible, to learn about the mechanisms generating observed patterns of spatial genetic variation. The first step for these investigations, and an important component for any study of IBE, is to carefully consider the potential processes that could be taking place. Below, we identify four ecological processes – not mutually exclusive – that can generate a pattern of IBE, including (i) natural selection against immigrants, (ii) sexual selection against immigrants, (iii) reduced hybrid fitness and (iv) biased dispersal (Fig. 2).

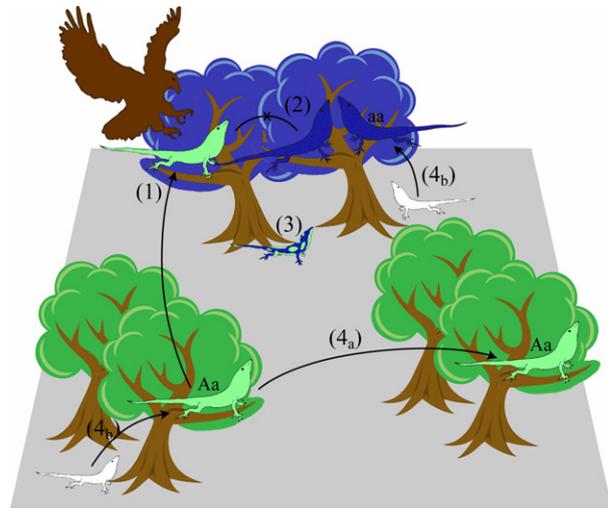


Fig. 2 Illustration of processes that can generate a pattern of isolation by environment. Dispersal between divergent environments can be reduced when (1) natural selection acts upon immigrants adapted to different environmental conditions, (2) sexual selection limits the reproductive success of immigrants with alternative traits, (3) hybrid offspring of native and immigrant parents have reduced fitness, for instance due to intermediate phenotypes, (4_a) biased dispersal resulting from a genotype or phenotype leads to a dispersal preference for particular environments or (4_b) biased dispersal resulting from a plastic natal habitat preference leads to a dispersal preference for similar habitats.

Natural selection against immigrants

Natural selection can generate IBE among populations inhabiting different environments when these populations are locally adapted (Nosil *et al.* 2005; Räsänen & Hendry 2008). In these cases, populations evolve traits suited to their local environments, regardless of their fitness consequences in other environments (Kawecki & Ebert 2004; Servedio 2004; Nosil *et al.* 2005). Thus, native genotypes in each environment will have, on average, higher fitness than immigrant genotypes originating in different environments (Kawecki & Ebert 2004; Servedio 2004). When individuals or populations show ecological specialization (Lu & Bernatchez 1999; Via & Hawthorne 2002), divergent natural selection will limit the reproductive success of individuals moving into different environments from which they are adapted (Räsänen & Hendry 2008; Mosca *et al.* 2012). For instance, walking sticks adapted to appear cryptic on certain host plants experience greater predation from visually oriented predators after moving to a different host species (Nosil 2004; Nosil *et al.* 2005), and divergent selection regimes in inland habitat with seasonal drought and coastal habitat with year round soil moisture cause nearly complete reductions in gene flow between populations of yellow monkey flowers adapted

to either climate (Lowry *et al.* 2008). We can expect the strength of divergent selection to be proportional to the magnitude of the differences among environments (Crispo *et al.* 2006; Lee & Mitchell-Olds 2011; Wang *et al.* 2013), and therefore, pairs of populations inhabiting increasingly different environments will experience reduced gene flow and greater genetic divergence (Barton 1979; Nosil *et al.* 2005; Thorpe *et al.* 2008). This environmentally associated natural selection may generate either pre- or postmating reproductive isolation, depending on whether immigrants survive and thrive long enough to mate locally. If immigrants are able to mate locally, there may subsequently be selection against immigrant alleles in hybrids (see '*Reduced Hybrid Fitness*' below).

Sexual selection against immigrants

Similarly, divergent sexual selection among populations inhabiting different environments can also generate IBE (Servedio 2004; Nosil *et al.* 2005; Safran *et al.* 2013). When populations inhabiting different environments show divergence in mate choice or sexual signals, sexual selection will reduce the reproductive success of dispersers moving between them (Servedio 2004; Nosil *et al.* 2005). In some cases, divergent sexual selection will be related to environmentally driven natural selection. For instance, under the good genes hypothesis, mate choice evolves so that individuals prefer mates possessing traits that increase offspring fitness (Ingleby *et al.* 2010), and if the fitness conferred by these traits varies across environments, divergent sexual selection will result and lead to variation in preferences for ecologically important traits (Nosil *et al.* 2005). This is the case with lesser wax moths, in which females choose males with different signals to produce offspring that mature faster in different food and temperature environments (Jia & Greenfield 1997). In other cases, divergent sexual selection can also be related to environmental variation but not natural selection. For instance, under the sensory drive hypothesis, sexual signals and their perception by receivers can evolve to be more effective under local environmental conditions, and therefore, dispersers may have reduced reproductive opportunities if their sexual signals are viewed in a different ecological context. Such is the case with some cichlids (Seehausen *et al.* 2008) and sticklebacks (Boughman 2001), in which adaptive variation in visual sensitivity leads to genetic isolation between groups expressing different nuptial coloration across gradients in ambient light conditions. Thus, divergent sexual selection can act in concert with divergent natural selection or independently of it (Servedio 2004; Nosil *et al.* 2005; Safran *et al.* 2013), in both cases producing

increased levels of IBE. As with natural selection against immigrants, imperfect sexual selection against nonlocal individuals can allow for the creation of hybrid offspring, the sexual characteristics and mating behaviour of which may themselves be selected against (see '*Reduced Hybrid Fitness*' below).

Reduced hybrid fitness

Selection can further contribute to IBE when hybrid offspring of immigrant-native parent crosses have reduced fitness relative to their nonhybrid neighbours (i.e. postzygotic extrinsic reproductive isolation of ecologically divergent populations; Nosil *et al.* 2005; McBride & Singer 2010). If hybrid offspring have intermediate phenotypes, then they may not occupy an available ecological niche in their natal environment or may have limited mating opportunities, both of which will reduce effective rates of long-term gene flow (Nosil *et al.* 2005; Garant *et al.* 2007; McBride & Singer 2010). For instance, in *Euphydryas* butterflies, hybrids from parents adapted to different hosts exhibit intermediate traits that are significantly maladaptive, including foraging and oviposition behaviours (McBride & Singer 2010). These cases will mostly serve to strengthen the patterns resulting from natural or sexual selection on immigrants. However, we consider this separate from these processes, as other authors have (e.g. Servedio 2004; Nosil *et al.* 2005), because selection acts on a different generation of individuals, rather than on the immigrants themselves, and under certain scenarios, the determinants of offspring fitness may be different from those of immigrant fitness. For example, if combinations of alleles that arose in isolated parental populations are incompatible, hybrid fitness may be reduced due to intrinsic, rather than extrinsic reproductive isolation (e.g. Dobzhansky-Muller incompatibilities; Dobzhansky 1937); in this case, selection would not be against immigrant alleles, but rather against the combination of immigrant and native alleles.

Additionally, when populations are separated by more than an individual's maximum dispersal distance, selection will act not against the immigrant individual but against immigrant alleles (i.e. through intervening populations over successive generations). With greater dispersal separation between populations comes more time over which recombination may break down linkage disequilibrium between neutral diversity and selected alleles on their journey from one environment to another. Larger distances between populations will therefore make it more likely that neutral diversity will escape its non-local background and be able to introgress into the local gene pool, decreasing a genome-wide pattern of IBE.

Biased dispersal

Isolation by environment can also result when a genotype, phenotype or behaviour contributes to a dispersal preference for a particular environment. Under these cases of 'biased' or 'directed' dispersal, an individual's traits affect the likelihood of moving to various habitats. This may be due to heritable variation (Edelaar *et al.* 2008; Bolnick & Otto 2013) or may be the result of a behavioural or plastic response induced by an individual's natal or developmental environment (Davis & Stamps 2004). A dispersal bias may arise because of a fitness or performance advantage in a particular environment, like the matching of coloration and habitat in white sand lizards (Rosenblum & Harmon 2011). In this case, the resulting pattern of dispersal naturally leads to the correlation of genotype and environment (Bolnick & Otto 2013). However, such a pattern may also arise even when individuals do not necessarily experience differential selection in different habitats, for instance when dispersers avoid novel habitat when moving across an environmentally heterogeneous landscape (Stevens *et al.* 2005; Feder & Forbes 2007) or when individuals prefer to disperse to habitats similar to their native habitat, known as natal habitat preference induction (Davis & Stamps 2004). This type of environmentally induced plastic response can be seen in two species of true frogs, in which the exposure of eggs to olfactory cues in water led to a preference among tadpoles for those environmental cues that is maintained through metamorphosis (Hepper & Waldman 1992). In such cases, there does not need to be local adaptation, but a pattern of IBE may still arise. We consider these processes distinct from natural or sexual selection against immigrants because selection may not actually act upon these individuals – there are no deaths or reproductive consequences because individuals move before any selective events can occur (Bolnick & Otto 2013).

Methods and considerations for studying IBE

Sampling scheme

When investigating patterns of spatial genetic variation, a researcher must design a sampling scheme that allows the study to disentangle the relative effects of geographic distance and ecological or environmental distance. This includes two important considerations: sampling to maximize the range of observed geographic and ecological distances and sampling to minimize the correlation between the potential explanatory variables. By sampling over an extensive range of geographic and environmental distances, the researcher can learn about the shape of the decay of genome-wide relatedness with

different distances. For example, if patterns of IBD or IBE increase nonlinearly, a researcher with a sampling scheme that includes only small distances and large distances might be unable to quantify well the shape of that curve (see Fig. 3).

Additionally, researchers must avoid a sampling scheme in which ecological and geographic distances are confounded. For example, in the confounded sampling configuration shown in Figure 3, geographic distance is strongly correlated with environmental distance, making it difficult to statistically disentangle their effects. Instead, samples from different environments should be drawn from a mixture of populations that are geographically close and distant, and samples from similar environments should be as well. This will serve to reduce the association between geographic and environmental distances.

For both considerations – maximizing the range of sampled pairwise distances and minimizing the correlation of covariates – a limited sampling scheme could lead to uninterpretable or misleading results. However, empirical researchers are frequently constrained in their sampling effort and are therefore forced to make compromises in the number of locations or individuals they are able to collect and genotype. When designing sampling schemes with these compromises in mind, we suggest first examining simple histograms of pairwise distances (geographic and environmental) and correlations between environmental and geographic distances under a projected sampling scheme prior to sample collection. If necessary, adjustments to the proposed scheme can then be made to increase the diversity of pairwise distances sampled or to minimize the correlation between these variables.

Above, we have placed an emphasis on placement and number of sampled locations, the latter of which is negatively correlated with the number of individuals per sampling location that a researcher can afford to genotype. Increasing the number of individuals sampled per location decreases the binomial sampling noise around estimates of population allele frequencies and is therefore useful in accurately estimating pairwise population differentiation. However, in the era of next-generation sequencing, it is possible to sequence each individual at large numbers of loci, which, assuming they are independent, serve as independent instantiations of the coalescent process. With many sequenced loci in two populations, it is therefore possible to get good estimates of the differentiation between them, even if the sample size at each locus is small, and the estimate of differentiation is therefore noisy (Patterson *et al.* 2006).

In summary, researchers designing studies to investigate IBE should seek to maximize the range of

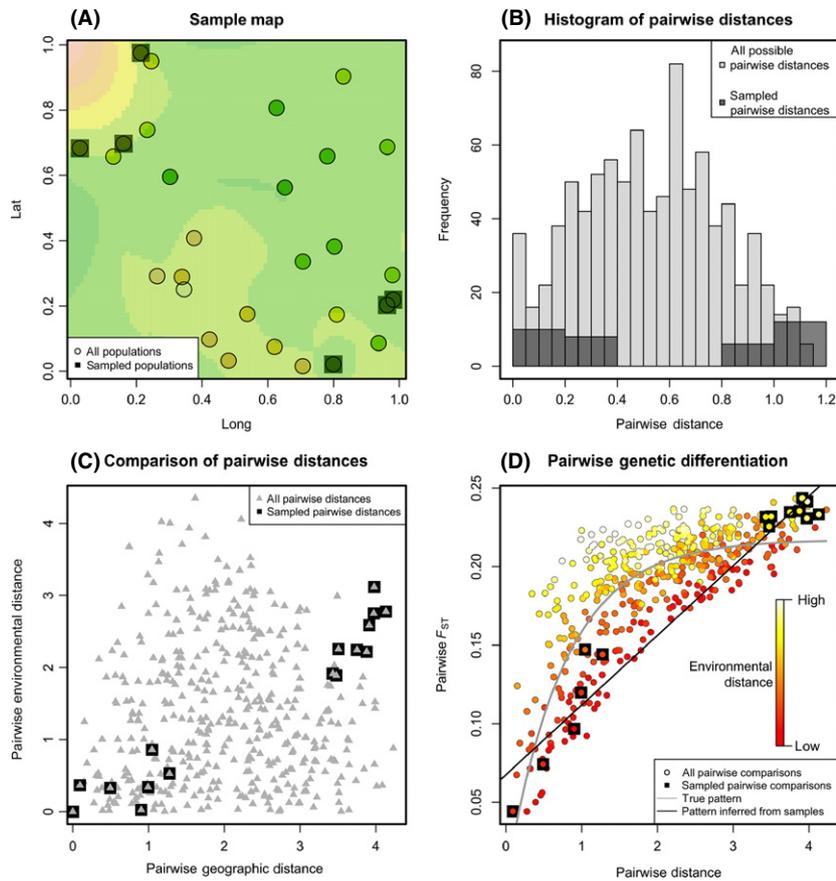


Fig. 3 Illustration of common pitfalls in designing a sampling scheme. These panels show the contrasted outcomes between complete sampling and an inadequately designed sampling scheme for populations on a hypothetical landscape with environmental variation (A). The histogram of pairwise distances (B) obtained by a poor sample design shows that this design results in obtaining pairwise distances that are not fully representative of the full set of populations. The comparison of the environmental and geographic distances recovered under the complete and partial sampling schemes (C) shows that these distances are highly correlated under the partial sampling scheme; IBD and IBE are conflated because all comparisons over short distances are also similar in environment, and all comparisons over long distances are environmentally disparate, with no intermediate comparisons to help disentangle the relative contributions of IBD and IBE. Finally, pairwise genetic differentiation plotted against pairwise geographic distance and coloured by environmental distance (D) shows that under the partial sampling scheme, the shapes of the curves for IBD and IBE are poorly estimated.

geographic and environmental distances between sampled locations, minimize the correlations of those geographic and environmental covariates and maximize the number of sampled locations even at the cost of sample size. In all cases, the relevant question that should be kept in mind is not whether it will be possible to detect spatial genetic differentiation but whether it will be possible to distinguish a genetic pattern of IBE from the background pattern of IBD.

Statistical techniques

This same question should be at the heart of the statistical techniques used to study IBE; researchers must account for IBD when attempting to quantify IBE along one or more environmental axes. This task is inherently difficult because measures of pairwise genetic distance are nonindependent and therefore should not be modelled directly as a function of pairwise geographic or ecological distances. The Mantel and partial Mantel tests were designed to avoid this problem of nonindependence by implementing a significance test that explicitly accounts for the pairwise nature of the dependent and independent variables. However, when the data are spatially autocorrelated, the partial Mantel has

a pathological type I error rate (Guillot & Rousset 2013), as the significance procedure implicitly rejects the spatial structure of the data.

However, the question the partial Mantel has been used to answer in landscape genetics is still vital to many research programmes today: What are the relative contributions to observed patterns of spatial genetic variation from geographic and ecological distances? Recently, several new, more statistically robust methods designed to answer this question have been released. These include methods for examining genome-wide patterns of differentiation and for investigating divergence in individual loci, both of which are important for understanding the nature of IBE. Coupled with new techniques for treating the spatial component of genetic variation more explicitly, these provide strength and flexibility for studying IBE in almost any natural system.

New methods for examining general patterns of genetic differentiation fall into two categories: modelling covariance in allele frequencies and matrix regression approaches. Of the former, BEDASSLE (Bradburd *et al.* 2013) is a Bayesian method that models the covariance in allele frequencies across the genome as a decreasing function of pairwise geographic and

ecological distances. The coefficients estimated for these pairwise distance elements can be compared to quantify the relative impact of geographic and ecological distances on patterns of genetic variation. This method was applied to a maize data set and revealed that 1000 m of elevation difference between populations has the same effect on genetic differentiation as about 150 km of horizontal distance (Bradburd *et al.* 2013). Of the latter, methods based on matrix regression, including generalized dissimilarity modelling (GDM; Freedman *et al.* 2010), structural equation modelling (SEM; Wang *et al.* 2013) and multiple matrix regression with randomization (MMRR; Wang 2013), apply a regression framework to simultaneously quantify the effects of multiple distance matrices on a single response variable, typically genetic distance, using different computational methodologies. SEM was used to infer that IBD typically contributed about twice as much to genetic divergence as IBE in 17 species of anoles and to quantify the relative contributions of individual environmental variables to IBE in those species (Wang *et al.* 2013). Both sets of methods have the power to disentangle IBD and IBE, and the preferred method to be used will depend upon the data available and the question to be answered.

New techniques for dealing with genetic distances in a spatially explicit manner include those that handle continuous spatial variation and those designed to account for the spatial arrangement of population networks. For continuous variation, LocalDiff (Duforet-Frebourg & Blum 2014) uses Bayesian kriging and MEMGENE uses Moran's eigenvector maps (Galpern *et al.* 2014) to detect discontinuities in patterns of gene flow across a landscape, which may be associated with landscape heterogeneity and barriers to dispersal. For population networks, the utilization of conditional genetic distances derived from population network topology can improve estimation of IBD and IBE and can be used with phylogeographic history to separate the effects of historical and contemporary barriers to gene flow (Dyer *et al.* 2010). This method was used to parse out the effects of phylogeographic history in the Sonoran desert succulent, revealing individual effects of spatial and bioclimatic variables on genetic differentiation (Dyer *et al.* 2010). Both these techniques are valuable for handling spatial genetic distance data, especially when paired with an appropriate statistical method for disentangling IBD and IBE.

These approaches all consider genome-wide patterns of differentiation and are therefore designed to answer a separate set of questions from methods such as Bayenv2 (Coop *et al.* 2010; Günther & Coop 2013), which asks whether individual SNPs are correlated with environmental variation, and spaMM (Rousset & Ferdy

2014), which is designed to account for spatial autocorrelation in the association of a genotype with some environmental variable. The former was applied to an Atlantic herring data set to identify loci that were strongly differentiated along a salinity gradient (Günther & Coop 2013). Once patterns of differentiation have been identified, additional methods can be applied to genomic data to attempt to elucidate the population-level processes that have generated those patterns. These methods, which are principally designed to look for the signature of selection across the whole genome, include identifying signals of selective sweeps and quantifying patterns of introgression in different regions of the genome characterized by different recombination rates (e.g. Geraldès *et al.* 2011; Nachman & Payseur 2012). All of these methods represent significant steps forward in the statistical analysis of patterns of spatial genetic variation and IBE; as next-generation sequencing power enables researchers to collect genomic data on ecological scales (many samples over broad geographic sampling areas), we expect both their use and their utility to increase.

Experimental data

Statistical techniques are valuable for quantifying IBE, but an experimental approach remains the best way to establish a causal mechanism. By manipulating organisms and the environments in which they occur, it is possible to distinguish between different processes that could produce IBE. A detailed review of the full range of experimental procedures used to learn about biological processes generating genetic variation is beyond the scope of this study. However, below, we highlight a selection of the most commonly used experimental tools and discuss the ways in which they may be used to learn about the processes generating a pattern of IBE.

To determine whether natural selection against immigrants is driving a pattern of IBE, researchers can use reciprocal transplant experiments, multiple common gardens or provenance tests (Thorpe *et al.* 2005; Leinonen *et al.* 2011). A fitness advantage observed when organisms are matched with their local environment is evidence for local adaptation and natural selection against immigrants. For instance, reciprocal transplant experiments on monkey flowers demonstrated natural selection against immigrants adapted to different soil moisture environments (Lowry *et al.* 2008).

To determine whether sexual selection has generated IBE, researchers can use mate choice trials. Individuals from the same environment mating assortatively when presented with options from similar and different environments can be taken as evidence that sexual selection is acting to shape patterns of genetic variation between

populations, as was the case with laboratory matings of cichlids that showed assortative mating between geographically close populations that differed in male nuptial coloration (Knight & Turner 2004).

Researchers studying organisms with sufficiently short generation times and tractable reproductive behaviour can create hybrids between populations from ecologically divergent habitats and assess their fitness in a common garden or in both parental environments to test for reduced hybrid fitness generating IBE. For instance, randomized common garden experiments on hybrid *Arabidopsis lyrata* from ecologically divergent populations with differences in timing of flowering and floral display traits demonstrated variation in the fitness of hybrids relative to parent populations (Leinonen *et al.* 2011).

Researchers should also consider the possibility that nonselective processes, such as biased dispersal or differentially resistant landscape elements, have led to decreased gene flow between populations. These possibilities can be experimentally examined using controlled or reciprocal release experiments (e.g. Bolnick *et al.* 2009), radio-tracking experiments (e.g. Broquet *et al.* 2006), stable isotope analysis (e.g. Pilot *et al.* 2012) or experimental quantification of an organism's dispersal ability in different environments (e.g. Stevens *et al.* 2005). In a controlled or reciprocal release, patterns of dispersal that are biased towards native habitat are evidence that observed IBE is due to biased dispersal. For example, in lake and stream sticklebacks, mark–transplant–recapture experiments showed that a large majority returned to their native habitat and that dispersal into non-native habitat was phenotype dependent (Bolnick *et al.* 2009). Radio-tracking and stable isotope analysis, which can reveal biased patterns of organisms moving over and utilizing different habitats within a landscape, can also provide strong evidence that biased dispersal is generating IBE. For instance, stable isotope profiles have been used to reveal correlations between diet differences, associated with habitat choice, and genetic distances in European wolves (Pilot *et al.* 2012). Finally, quantification of an organisms' abilities to disperse over or between different substrates can also provide indicators of biased dispersal. For instance, mobility analysis in an experimental arena composed of different habitats provided quantitative estimates of dispersal ability over different substrates in the natterjack toad (Stevens *et al.* 2005).

Finally, because these processes are not mutually exclusive, multiple processes may act or have acted to generate the observed pattern of IBE. For example, with reinforcement, the presence of reduced hybrid fitness is expected in the long run to select for increased pre-mating sexual reproductive isolation between parental pop-

ulations. Therefore, it may be necessary to follow multiple lines of experimental evidence to determine the ecological processes that have generated patterns of spatial genetic differentiation.

Caveats

There are a number of factors that can potentially confound the detection and measurement of IBE. Population history, demography and heterogeneity may all influence estimates of observed IBE and potentially lead to migration-drift disequilibrium. Sampling design can also compound the challenges posed by these factors, potentially leading to inferential errors. If, for example, some sampled populations are only recently diverged and not in migration-drift equilibrium, the estimated rates of IBD and IBE in the complete set of populations may be considerably noisier (Slatkin 1993; Marko & Hart 2011). In addition, model inadequacy poses a serious problem in the analysis of landscape genetic data, in which the processes that have generated the data are almost sure to be vastly more complex and idiosyncratic than the models used to perform inference. Researchers are well advised to assess model adequacy, either through evaluating model fit or by performing an explicit test of adequacy, such as posterior predictive simulation (e.g. Bradburd *et al.* 2013), and to be cautious in the interpretation of their results. Recognizing these potentially confounding factors and properly designing a study to account for them is critical for accurately detecting IBE.

Broader implications

The study of IBE and the mechanisms generating it have significant implications for a variety of disciplines. The most obvious example is landscape genetics – much of landscape genetics has focused on examining how landscapes influence population connectivity (Storfer *et al.* 2007; Sork & Waits 2010; Wagner & Fortin 2013), and analysis of IBE is clearly important for fully understanding how landscape and environmental features influence gene flow and population structure (Bradburd *et al.* 2013; Wang *et al.* 2013; Sexton *et al.* 2014). This can add a crucial element for assessing the importance of different parts of the landscape for corridor and reserve design and for performing long-term population viability analyses, which are clearly valuable for conservation efforts. For species in which IBE is prominent, conservationists might also have to consider whether IBE results from local adaptation and whether to prioritize the protection of gene flow between more similar

habitats over gene flow from divergent habitats that could 'swamp out' locally adaptive genotypes (Aitken & Whitlock 2013).

Along these lines, IBE can also contribute to studying the ecology of local adaptation, currently a topic of major interest (Aitken & Whitlock 2013; Blanquart *et al.* 2013; Günther & Coop 2013; Butlin *et al.* 2014), potentially providing an efficient first step in identifying systems with adaptive divergence. However, because many mechanisms can generate IBE, detection of this pattern alone is not evidence of local adaptation. Similarly, under the simplifying assumption that IBE always results from selection, some previous studies have interpreted IBE as evidence for incipient ecological speciation (Shafer & Wolf 2013). By definition, ecological speciation is the evolution of reproductive isolating barriers due to divergent selection under different ecological conditions (Lu & Bernatchez 1999; Schluter 2009; Thibert-Plante & Hendry 2010). Because IBE can result from processes other than selection, it should not be taken at face value as evidence for ecological speciation; a more prudent interpretation is that IBE can indicate that the underlying conditions necessary for ecological speciation may exist in a given system. Nevertheless, IBE is still valuable for studying local adaptation and ecological speciation because many methods can evaluate which environmental variables contribute to IBE, and this can aid in designing experiments to identify the ecological factors driving adaptive population divergence and isolation. Moreover, because of its association with gene flow, IBE is also valuable for examining the important question of how local adaptation occurs in the face of ongoing gene flow (Saint-Laurent *et al.* 2003; Nosil & Crespi 2004; Räsänen & Hendry 2008; Butlin *et al.* 2014; Muir *et al.* 2014). In these scenarios, the pattern of gene flow may be more important than the level of overall gene flow – for instance if gene flow is primarily among similar environments – and studies that contrast IBE with IBD could contribute to our understanding of the types of gene flow that enable adaptive population divergence.

Finally, studies of IBE can also influence landscape and community ecology. For instance, the general prevalence of IBE in nature (Shafer & Wolf 2013; Sexton *et al.* 2014) has ramifications for traditional ecological theory about the distribution of individuals in space, like the ideal free distribution, which posits that organisms are free to move among habitat patches unimpeded and will distribute themselves among them in proportion to the availability of resources. In general, a pattern of IBE means that these assumptions are unmet, as individuals will either have a nonrandom distribution among patches driven by factors aside from resource availability (Nosil *et al.* 2005; Bolnick & Otto

2013) or will be unable to move freely among patches because of some spatial ecological process mediating dispersal (Lu & Bernatchez 1999; Crispo *et al.* 2006; Räsänen & Hendry 2008; Bradburd *et al.* 2013). Additionally, if IBE is present across many species within a community, then they may show spatially similar patterns of divergence among populations, and this could potentially lead to the codiversification of multiple species (Johnson & Stinchcombe 2007). Thus, IBE, its prevalence and its underlying factors can have significant implications for a wide variety of ecological and evolutionary processes.

Future directions

Despite the growing interest in IBE, many exciting areas remain open for future research (Balkenhol *et al.* 2009; Storfer *et al.* 2010). Here, we outline five areas of pressing interest that present a wealth of opportunities for innovative research in the near future: (i) landscape genomics, (ii) comparative landscape genetics, (iii) population heterogeneity, (iv) temporal variation and (5) identifying the underlying ecological processes that drive IBE. Investigating these areas and answering the important questions they present will greatly expand our knowledge of how ecology influences genetic variation across space, time, taxa and the genome.

Landscape genomics

Integrating population genomics into landscape ecological research is an exciting frontier for landscape genetics that will open up many new avenues of scientific inquiry. Several recent studies have already explored how landscape genomics can provide greater power and resolution for examining spatial patterns and adaptive variation (e.g. Coop *et al.* 2010; Lasky *et al.* 2012; Parchman *et al.* 2012; Vincent *et al.* 2013; Yoder *et al.* 2014). However, one question that has not yet been extensively investigated is how the environment influences variation differentially across the genome.

Different sites across the genome can experience different evolutionary scenarios because of the dynamics of the many processes that act on genetic variation (Turner & Hahn 2010; Flaxman *et al.* 2013; Soria-Carrasco *et al.* 2014). While the neutral process of drift due to decreased gene flow between a pair of populations acts on the entire genome, selective forces, such as natural and sexual selection against immigrants, will only target the loci involved in the traits under selection (Charlesworth *et al.* 1997; Turner & Hahn 2010). A pattern of IBE due to selective forces may therefore be observed locally at a given locus but not be seen globally across the entire genome, and the localization of

these effects will depend partly on the rate of recombination, which itself may vary across the genome (Nachman & Payseur 2012). Recent advances in population genetics have significantly improved the inference of heterogeneous coalescent histories across the genome (Harris & Nielsen 2013; Ralph & Coop 2013), and we think that the incorporation of ecological processes into these methods will be an exciting way forward. This could reveal how various signatures of IBE due to different underlying environmental factors are expressed across the genome – with regard to the localization and relative strengths of IBD and IBE across gene regions with different functions, architecture and recombination rates – and such studies could yield unprecedented looks into the ecological factors that drive genetic divergence in nature.

Comparative landscape genetics

Studies of single species on single landscapes are undoubtedly valuable for landscape genetics, often providing important information on organisms that are ecologically interesting or of conservation concern. However, comparative studies, either of multiple species or of multiple landscapes, are probably to provide the next big advances for understanding organism–landscape interactions. These comparisons can reveal the factors that drive IBE, generally, and whether they are intrinsic to organisms or to landscapes. Examinations of multiple species on one landscape (e.g. Goldberg & Waits 2010; Richardson 2012) can answer whether multiple taxa are affected in similar ways by a particular landscape, and studies of single species across multiple landscapes (e.g. Short Bull *et al.* 2011; Trumbo *et al.* 2013) can answer whether the influences of landscapes on organisms are the result of the specific spatial structure of the landscape or the biology of the organisms themselves. These studies may be of particular interest when trophic relationships between study organisms are known and provide a set of predictions about their relative patterns of population structure (e.g. patterns of structure in predator–prey or host–parasite systems), but all such studies will help in understanding how organism–landscape interactions contribute to IBE.

Population heterogeneity

To date, most studies have performed their analyses at the species or meta-population level, assuming that ecological responses are essentially constant across all populations. However, populations may diverge in ways that are important for how they interact with the landscape, including dispersal ability, habitat preference and adaptation to different ecological conditions, all of

which may influence patterns of gene flow. Spatial variation in evolutionary processes, such as selection, can also affect gene flow and lead to different factors driving IBE in different populations. So, despite a focus on spatial variation, few landscape genetic studies have considered how spatial variation in the traits and factors affecting gene flow contributes to estimates of IBE and its causal factors at higher levels. If populations vary substantially in their disposition towards IBD and IBE, then the subset of populations chosen for an empirical study may also influence the overall estimation of landscape effects on a given species. In any case, new studies that explicitly consider this possibility will provide insight into how spatial variation among populations shapes IBE.

Temporal variation

Another important consideration for studies of IBE is temporal variation, especially because evolutionary forces, ecological processes and environments typically change through time. While spatial variation and scale have been identified as important factors in landscape genetic analysis (Cushman & Landguth 2010), the implications of temporal variation have not yet been thoroughly investigated, although some recent studies provide indications of their importance (Crispo & Chapman 2009; Dyer *et al.* 2010; Epps *et al.* 2013; He *et al.* 2013). Ideally, studies would be conducted on temporally spaced samples and geospatial data from corresponding time periods, potentially including museum specimens (Nachman 2013), but new analytical methods may also allow inferences of historical patterns (Dyer *et al.* 2010; He *et al.* 2013). In either case, these studies should provide valuable insights into the tempo of change in ecological drivers of diversification and the relative importance of historical landscape factors for explaining contemporary patterns of variation. This is particularly important for understanding how species will respond to changing climate and environmental conditions, a commonly stated goal of landscape genetic studies. Probably the best way to predict how species will be affected by a changing environment is to account for how they responded to changing conditions before.

Identifying underlying ecological processes

Finally, while the identification of IBD, IBE and related patterns still presents many interesting research objectives, we believe that it is important for future research to go beyond describing patterns of spatial variation and to begin testing for the underlying mechanisms that generate these patterns. One of the fundamental

charges of ecology and evolutionary biology is to explain the patterns of variation – both phenotypic and genetic – that we observe in nature. The first step is quantifying the observed patterns, and the second is investigating the processes that generate them. Sometimes this may mean that research on IBE will have to extend beyond landscape genetics techniques and use experimental and traditional landscape ecological methods, such as reciprocal transplants, common gardens, radio-tracking, functional morphology and biomechanics, to examine whether divergent populations exhibit differences consistent with a particular mode of differentiation. Targeted population genetic analyses, applied with proper sampling designs, can also be valuable for examining spatial variation in signals of selection and the dispersion of genotypes linked to ecologically important traits across environmental gradients. These studies will significantly expand the scope of investigations into IBE and should provide remarkable new insights into the causal ecological factors driving genetic variation in the wild.

Conclusions

Over the last decade, landscape genetics has made remarkable progress in examining the impacts of landscape variation on gene flow and population dynamics (Sork & Waits 2010; Storfer *et al.* 2010). Isolation by environment will play a major part in the next big steps, providing a framework for examining how ecological and environmental heterogeneity shape the distribution of genetic variation in nature. The environment is clearly a core component of the landscape – even though it has not been explicitly considered in landscape genetics until recently – which can significantly influence gene flow and population connectivity. As suggested by a handful of recent studies, this effect could be widespread (Shafer & Wolf 2013; Sexton *et al.* 2014), although more detailed studies of IBE are now necessary to determine its full extent, the relative strengths of the various processes that underlie it and the range of ecological conditions necessary for its generation.

Isolation by environment is distinct from isolation by distance, which has been the foundation for examining gene flow and population structure for most of a century. Hence, the study of IBE will provide new insights into mechanisms of dispersal, patterns of connectivity and modes of differentiation both within and between species. These will likely have major implications for a wide variety of disciplines, and they should be especially important for understanding how organisms will respond to rapid ecological change. By incorporating environmental forces and ecological interactions – and potentially their spatial variation – into

analyses of genetic divergence, studies of IBE present opportunities to identify the important factors that influence population, community and even ecosystem dynamics in space. Thus, the conceptual framework of IBE will play an important role in advancing our understanding of how ecology shapes the evolution of biological diversity.

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References

- Aitken SN, Whitlock MC (2013) Assisted gene flow to facilitate local adaptation to climate change. *Annual Review of Ecology, Evolution, and Systematics*, **44**, 367–388.
- Andrew RL, Ostevik KL, Ebert DP, Rieseberg LH (2012) Adaptation with gene flow across the landscape in a dune sunflower. *Molecular Ecology*, **21**, 2078–2091.
- Balkenhol N, Gugerli F, Cushman S *et al.* (2009) Identifying future research needs in landscape genetics: where to from here? *Landscape Ecology*, **24**, 455–463.
- Barton NH (1979) Gene flow past a cline. *Heredity*, **43**, 333–339.
- Blanquart F, Kaltz O, Nuismer SL, Gandon S (2013) A practical guide to measuring local adaptation. *Ecology Letters*, **16**, 1195–1205.
- Bolnick DI, Otto SP (2013) The magnitude of local adaptation under genotype-dependent dispersal. *Ecology and Evolution*, **3**, 4722–4735.
- Bolnick DI, Snowberg LK, Patenia C *et al.* (2009) Phenotype-dependent native habitat preference facilitates divergence between parapatric lake and stream stickleback. *Evolution; International Journal of Organic Evolution*, **63**, 2004–2016.
- Boughman JW (2001) Divergent sexual selection enhances reproductive isolation in sticklebacks. *Nature*, **411**, 944–948.
- Bradburd GS, Ralph PL, Coop GM (2013) Disentangling the effects of geographic and ecological isolation on genetic differentiation. *Evolution*, **67**, 3258–3273.
- Broquet T, Johnson CA, Petit E *et al.* (2006) Dispersal and genetic structure in the American marten, *Martes americana*. *Molecular ecology*, **15**, 1689–1697.
- Butlin RK, Saura M, Charrier G *et al.* (2014) Parallel evolution of local adaptation and reproductive isolation in the face of gene flow. *Evolution*, **68**, 935–949.
- Charlesworth B, Nordborg M, Charlesworth D (1997) The effects of local selection, balanced polymorphism and background selection on equilibrium patterns of genetic diversity in subdivided populations. *Genetical Research*, **70**, 155–174.
- Claremont M, Williams ST, Barraclough TG, Reid DG (2011) The geographic scale of speciation in a marine snail with high dispersal potential. *Journal of Biogeography*, **38**, 1016–1032.
- Coop G, Witonsky D, Di Rienzo A, Pritchard JK (2010) Using environmental correlations to identify loci underlying local adaptation. *Genetics*, **185**, 1411–1423.

- Crispo E, Chapman LJ (2009) Temporal variation in population genetic structure of a Riverine African Cichlid fish. *Journal of Heredity*, **101**, 97–106.
- Crispo E, Bentzen P, Reznick DN, Kinnison MT, Hendry AP (2006) The relative influence of natural selection and geography on gene flow in guppies. *Molecular Ecology*, **15**, 49–62.
- Cushman S, Landguth E (2010) Scale dependent inference in landscape genetics. *Landscape Ecology*, **25**, 967–979.
- Davis JM, Stamps JA (2004) The effect of natal experience on habitat preferences. *Trends in Ecology & Evolution*, **19**, 411–416.
- Dobzhansky TG (1937) *Genetics and the Origin of Species*. Columbia University Press, New York.
- Duforet-Frebourg N, Blum MGB (2014) Nonstationary patterns of isolation-by-distance: inferring measures of local genetic differentiation with Bayesian kriging. *Evolution*, **68**, 1110–1123.
- Dyer RJ, Nason JD, Garrick RC (2010) Landscape modelling of gene flow: improved power using conditional genetic distance derived from the topology of population networks. *Molecular Ecology*, **19**, 3746–3759.
- Edelaar P, Siepielski AM, Clobert J (2008) Matching habitat choice causes directed gene flow: a neglected dimension in evolution and ecology. *Evolution*, **62**, 2462–2472.
- Edelaar P, Alonso D, Lagerveld S, Senar JC, Björklund M (2012) Population differentiation and restricted gene flow in Spanish crossbills: not isolation-by-distance but isolation-by-ecology. *Journal of Evolutionary Biology*, **25**, 417–430.
- Epps CW, Wasser SK, Keim JL, Mutayoba BM, Brashares JS (2013) Quantifying past and present connectivity illuminates a rapidly changing landscape for the African elephant. *Molecular Ecology*, **22**, 1574–1588.
- Feder JL, Forbes AA (2007) Habitat avoidance and speciation for phytophagous insect specialists. *Functional Ecology*, **21**, 585–597.
- Feder JL, Nosil P (2010) The efficacy of divergence hitchhiking in generating genomic islands during ecological speciation. *Evolution*, **64**, 1729–1747.
- Flaxman SM, Feder JL, Nosil P (2013) Genetic hitchhiking and the dynamic buildup of genomic divergence during speciation with gene flow. *Evolution*, **67**, 2577–2591.
- Foll M, Gaggiotti O (2006) Identifying the environmental factors that determine the genetic structure of populations. *Genetics*, **174**, 875–891.
- Freedman AH, Thomassen HA, Buermann W, Smith TB (2010) Genomic signals of diversification along ecological gradients in a tropical lizard. *Molecular Ecology*, **19**, 3773–3788.
- Galpern P, Peres-Nato PR, Polfus J, Manseau M (2014) MEMGENE: Spatial pattern detection in genetic distance data. *Methods in Ecology and Evolution*, **5**, 2001–2005.
- Garant D, Forde SE, Hendry AP (2007) The multifarious effects of dispersal and gene flow on contemporary adaptation. *Functional Ecology*, **21**, 434–443.
- Geraldes A, Basset P, Smith KL, Nachman MW (2011) Higher differentiation among subspecies of the house mouse (*Mus musculus*) in genomic regions with low recombination. *Molecular ecology*, **20**, 4722–4736.
- Goldberg CS, Waits LP (2010) Comparative landscape genetics of two pond-breeding amphibian species in a highly modified agricultural landscape. *Molecular Ecology*, **19**, 3650–3663.
- Guillot G, Rousset F (2013) Dismantling the Mantel tests. *Methods in Ecology and Evolution*, **4**, 336–344.
- Günther T, Coop G (2013) Robust identification of local adaptation from allele frequencies. *Genetics*, **195**, 205–220.
- Harris K, Nielsen R (2013) Inferring demographic history from a spectrum of shared haplotype lengths. *PLoS Genetics*, **9**, e1003521.
- He Q, Edwards DL, Knowles LL (2013) Integrative testing of how environments from the past to the present shape genetic structure across landscapes. *Evolution*, **67**, 3386–3402.
- Hepper PG, Waldman B (1992) Embryonic olfactory learning in frogs. *The Quarterly Journal of Experimental Psychology Section B*, **44**, 179–197.
- Hopkins R, Levin DA, Rausher MD (2012) Molecular signatures of selection on reproductive character displacement of flower color in *Phlox drummondii*. *Evolution*, **66**, 469–485.
- Ingleby FC, Hunt J, Hosken DJ (2010) The role of genotype-by-environment interactions in sexual selection. *Journal of Evolutionary Biology*, **23**, 2031–2045.
- Jia F-Y, Greenfield MD (1997) When are good genes good? Variable outcomes of female choice in wax moths *Proceedings of the Royal Society of London. Series B: Biological Sciences*, **264**, 1057–1063.
- Johnson MTJ, Stinchcombe JR (2007) An emerging synthesis between community ecology and evolutionary biology. *Trends in Ecology & Evolution*, **22**, 250–257.
- Kawecki TJ, Ebert D (2004) Conceptual issues in local adaptation. *Ecology Letters*, **7**, 1225–1241.
- Knight ME, Turner GF (2004) Laboratory mating trials indicate incipient speciation by sexual selection among populations of the cichlid fish *Pseudotropheus zebra* from Lake Malawi. *Proceedings of the Royal Society of London. Series B: Biological Sciences*, **271**, 675–680.
- Lasky JR, Des Marais DL, McKay JK *et al.* (2012) Characterizing genomic variation of *Arabidopsis thaliana*: the roles of geography and climate. *Molecular Ecology*, **21**, 5512–5529.
- Lee C-R, Mitchell-Olds T (2011) Quantifying effects of environmental and geographical factors on patterns of genetic differentiation. *Molecular Ecology*, **20**, 4631–4642.
- Leinonen PH, Remington DL, Savolainen O (2011) Local adaptation, phenotypic differentiation, and hybrid fitness in diverged natural populations of *Arabidopsis lyrata*. *Evolution*, **65**, 90–107.
- Lowry DB, Rockwood RC, Willis JH (2008) Ecological reproductive isolation of coast and inland races of *mimulus guttatus*. *Evolution*, **62**, 2196–2214.
- Lu G, Bernatchez L (1999) Correlated trophic specialization and genetic divergence in sympatric lake whitefish ecotypes (*Coregonus clupeaformis*): support for the ecological speciation hypothesis. *Evolution*, **53**, 1491–1505.
- Marko PB, Hart MW (2011) The complex analytical landscape of gene flow inference. *Trends in Ecology & Evolution*, **26**, 448–456.
- Mayr E (1963) *Animal Species and Evolution*. Harvard University Press, Cambridge, MA.
- McBride CS, Singer MC (2010) Field studies reveal strong post-mating isolation between ecologically divergent butterfly populations. *PLoS Biology*, **8**, e1000529.
- McRae BH, Beier P (2007) Circuit theory predicts gene flow in plant and animal populations. *Proceedings of the National Academy of Sciences*, **104**, 19885–19890.
- Meirns PG (2012) The trouble with isolation by distance. *Molecular Ecology*, **21**, 2839–2846.

- Mosca E, Eckert AJ, Di Piero EA *et al.* (2012) The geographical and environmental determinants of genetic diversity for four alpine conifers of the European Alps. *Molecular Ecology*, **21**, 5530–5545.
- Muir AP, Biek R, Thomas R, Mable BK (2014) Local adaptation with high gene flow: temperature parameters drive adaptation to altitude in the common frog (*Rana temporaria*). *Molecular Ecology*, **23**, 561–574.
- Murphy MA, Dezzani R, Pilliod DS, Storfer A (2010) Landscape genetics of high mountain frog metapopulations. *Molecular Ecology*, **19**, 3634–3649.
- Nachman MW (2013) Genomics and museum specimens. *Molecular Ecology*, **22**, 5966–5968.
- Nachman MW, Payseur BA (2012) Recombination rate variation and speciation: theoretical predictions and empirical results from rabbits and mice. *Philosophical Transactions of the Royal Society, Series B*, **367**, 409–421.
- Nosil P (2004) Reproductive isolation caused by visual predation on migrants between divergent environments. *Proceedings of the Royal Society of London. Series B: Biological Sciences*, **271**, 1521–1528.
- Nosil P, Crespi BJ (2004) Does gene flow constrain adaptive divergence or vice versa? A test using ecomorphology and sexual isolation in *Timema cristinae* walking-sticks. *Evolution*, **58**, 102–112.
- Nosil P, Vines TH, Funk DJ (2005) Reproductive isolation caused by natural selection against immigrants from divergent habitats. *Evolution*, **59**, 705–719.
- Nosil P, Egan SP, Funk DJ (2008) Heterogeneous genomic differentiation between walking-stick ecotypes: “isolation by adaptation” and multiple roles for divergent selection. *Evolution*, **62**, 316–336.
- Parchman TL, Gompert Z, Mudge J *et al.* (2012) Genome-wide association genetics of an adaptive trait in lodgepole pine. *Molecular Ecology*, **21**, 2991–3005.
- Patterson N, Price AL, Reich D (2006) Population structure and eigenanalysis. *PLoS Genetics*, **2**, e190.
- Pilot M, Jędrzejewski W, Sidorovich VE, Meier-Augenstein W, Hoelzel AR (2012) Dietary differentiation and the evolution of population genetic structure in a highly mobile carnivore. *PLoS ONE*, **7**, e39341.
- Ralph P, Coop G (2013) The geography of recent genetic ancestry across Europe. *PLoS Biology*, **11**, e1001555.
- Räsänen K, Hendry AP (2008) Disentangling interactions between adaptive divergence and gene flow when ecology drives diversification. *Ecology Letters*, **11**, 624–636.
- Richardson JL (2012) Divergent landscape effects on population connectivity in two co-occurring amphibian species. *Molecular Ecology*, **18**, 4437–4451.
- Rosenblum EB, Harmon LJ (2011) “Same same but different”: replicated ecological speciation at white sands. *Evolution*, **65**, 946–960.
- Rousset F (1997) Genetic differentiation and estimation of gene flow from F-statistics under isolation by distance. *Genetics*, **145**, 1219–1228.
- Rousset F, Ferdy J-B (2014) Testing environmental and genetic effects in the presence of spatial autocorrelation. *Ecography*, **37**, 781–790.
- Safran RJ, Scordato ESC, Symes LB, Rodríguez RL, Mendelson TC (2013) Contributions of natural and sexual selection to the evolution of premating reproductive isolation: a research agenda. *Trends in Ecology & Evolution*, **28**, 643–650.
- Saint-Laurent R, Legault M, Bernatchez L (2003) Divergent selection maintains adaptive differentiation despite high gene flow between sympatric rainbow smelt ecotypes (*Osmerus mordax* Mitchell). *Molecular Ecology*, **12**, 315–330.
- Schluter D (2009) Evidence for ecological speciation and its alternative. *Science*, **323**, 737–741.
- Seehausen O, Terai Y, Magalhaes IS *et al.* (2008) Speciation through sensory drive in cichlid fish. *Nature*, **455**, 620–626.
- Servedio MR (2004) The evolution of premating isolation: local adaptation and natural and sexual selection against hybrids. *Evolution*, **58**, 913–924.
- Sexton JP, Hangartner SB, Hoffmann AA (2014) Genetic isolation by environment or distance: which pattern of gene flow is most common? *Evolution*, **68**, 1–15.
- Shafer ABA, Wolf JBW (2013) Widespread evidence for incipient ecological speciation: a meta-analysis of isolation-by-ecology. *Ecology Letters*, **16**, 940–950.
- Short Bull RA, Cushman SA, Mace R *et al.* (2011) Why replication is important in landscape genetics: American black bear in the Rocky Mountains. *Molecular Ecology*, **20**, 1092–1107.
- Slatkin M (1993) Isolation by distance in equilibrium and non-equilibrium populations. *Evolution*, **47**, 264–279.
- Soria-Carrasco V, Gompert Z, Comeault AA *et al.* (2014) Stick insect genomes reveal natural selection’s role in parallel speciation. *Science*, **344**, 738–742.
- Sork VL, Waits L (2010) Contributions of landscape genetics – approaches, insights, and future potential. *Molecular Ecology*, **19**, 3489–3495.
- Stevens VM, Polus E, Wesselingh RA, Schtickzelle N, Baguette M (2005) Quantifying functional connectivity: experimental evidence for patch-specific resistance in the Natterjack toad (*Bufo calamita*). *Landscape Ecology*, **19**, 829–842.
- Storfer A, Murphy MA, Evans JS *et al.* (2007) Putting the “landscape” in landscape genetics. *Heredity*, **98**, 128–142.
- Storfer A, Murphy MA, Spear SF, Holderegger R, Waits LP (2010) Landscape genetics: where are we now? *Molecular Ecology*, **19**, 3496–3514.
- Thibert-Plante X, Hendry AP (2010) When can ecological speciation be detected with neutral loci? *Molecular Ecology*, **19**, 2301–2314.
- Thorpe RS, Reardon JT, Malhotra A (2005) Common garden and natural selection experiments support ecotypic differentiation in the Dominican anole (*Anolis oculatus*). *The American Naturalist*, **165**, 495–504.
- Thorpe RS, Surget-Groba Y, Johansson H (2008) The relative importance of ecology and geographic isolation for speciation in anoles. *Philosophical Transactions of the Royal Society B: Biological Sciences*, **363**, 3071–3081.
- Trumbo DR, Spear SF, Baumsteiger J, Storfer A (2013) Range-wide landscape genetics of an endemic Pacific northwestern salamander. *Molecular Ecology*, **22**, 1250–1266.
- Turner TL, Hahn MW (2010) Genomic islands of speciation or genomic islands and speciation? *Molecular Ecology*, **19**, 848–850.
- Via S, Hawthorne DJ (2002) The genetic architecture of ecological specialization: correlated gene effects on host use and

- habitat choice in pea aphids. *The American Naturalist*, **159**, S76–S88.
- Vincent B, Dionne M, Kent MP, Lien S, Bernatchez L (2013) Landscape genomics in Atlantic Salmon (*salmo Salar*): searching for gene-environment interactions driving local adaptation. *Evolution*, **67**, 3469–3487.
- Wagner HH, Fortin M-J (2013) A conceptual framework for the spatial analysis of landscape genetic data. *Conservation Genetics*, **14**, 253–261.
- Wang IJ (2012) Environmental and topographic variables shape genetic structure and effective population sizes in the endangered Yosemite toad. *Diversity and Distributions*, **18**, 1033–1041.
- Wang IJ (2013) Examining the full effects of landscape heterogeneity on spatial genetic variation: a multiple matrix regression approach for quantifying geographic and ecological isolation. *Evolution*, **16**, 175–182.
- Wang IJ, Summers K (2010) Genetic structure is correlated with phenotypic divergence rather than geographic isolation in the highly polymorphic strawberry poison-dart frog. *Molecular Ecology*, **19**, 447–458.
- Wang IJ, Glor RE, Losos JB (2013) Quantifying the roles of ecology and geography in spatial genetic divergence. *Ecology Letters*, **16**, 175–182.
- Wright S (1943) Isolation by distance. *Genetics*, **28**, 114–138.
- Yoder JB, Stanton-Geddes J, Zhou P *et al.* (2014) Genomic signature of adaptation to climate in *Medicago truncatula*. *Genetics*, **96**, 1263–1275.

The authors developed the ideas for and wrote the article together.

Supporting information

Additional supporting information may be found in the online version of this article.

Fig. S1 Animated 3D plot of isolation by distance and environment. This is an 'animated gif' that rotates to show a 3D plot of a simulated data set in which both IBD and IBE are clearly seen. Points represent the genetic distance between a pair of populations plotted against their geographic and environmental distances and are heat-coloured by the magnitude of that environmental distance.